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Syntheses of *N*-Acyl and *N*-Alkoxycarbonyl Derivatives of 2-

Alkoxycarbonylamino-2-deoxy-D-glucose Dominique Lafont^a; Paul Boullanger^a

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SYNTHESES OF N-ACYL AND N-ALKOXYCARBONYL DERIVATIVES

OF 2-ALKOXYCARBONYLAMINO-2-DEOXY-D-GLUCOSE

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ABSTRACT

N-Acyl and *N*-alkoxycarbonyl derivatives of 1,3,4,6-tetra-*O*-acetyl-2-alkoxycarbonylamino-2-deoxy- β -D-glucopyranose have been synthesized using mixed anhydrides and symmetrical or disymetrical pyrocarbonates. These derivatives have been used as donors in 1,2-*trans*-glycosylation reactions promoted by Lewis acids. Besides the expected β -D-glycosides, cyclisation and rearrangement side-products were often encountered in such glycosylations.

INTRODUCTION

The high reactivity of 1,3,4,6-tetra-O-acetyl-2-allyloxycarbonylamino-2-deoxy- β -D-glucopyranose **1** as donor in glycosylation reactions has been demonstrated using various alcohols.¹⁻⁴ In order to extend the field of application of the method to donors disubstituted on nitrogen, we prepared *N*-acyl-*N*-alkoxycarbonyl and *N*,*N*-dialkoxycarbonyl analogues of compound **1**.

To our knowledge, other than the well-known phthalimido derivatives,⁵ only a few examples of N,N-diacyl derivatives of D-glucosamine have been reported in the literature. These compounds have been previously prepared by N-acetylation or N-benzoylation of 2-acetamido and 2-benzamido-D-glucose derivatives.⁶⁻⁸

RESULTS AND DISCUSSION

U. Ragnarsson and co-workers have recently described a general method for exhaustive *tert*-butoxycarbonylation of amides⁹ and carbamates¹⁰ under very mild conditions, by treatment with di-*tert*-butyl pyrocarbonate and a catalytic amount of 4-dimethylaminopyridine in dry acetonitrile. Similar protection of the amine function (N,N-dialkoxycarbonyl derivative) was reported earlier by F. M. F. Chen and N. L. Benoiton¹¹ in the field of aminoacids.

Application of the *N-tert*-butoxycarbonylation method to 1,3,4,6-tetra-*O*-acetyl-2allyloxycarbonylamino-2-deoxy- β -D-glucopyranose **1** and to 2-acetamido-1,3,4,6-tetra-*O*acetyl-2-deoxy- β -D-glucopyranose **2** afforded compounds **3** and **4** in high yields (87 % and 85 %, respectively).

However, as we reported earlier,¹² the *N*-tert-butoxycarbonyl group was not stable enough to be used as a protecting group in glycosylation reactions catalyzed by Lewis acids. Therefore, we tried to prepare N_*N -diallyloxycarbonyl derivatives of D-glucosamine. Attempts to use the commercially available diallyl pyrocarbonate¹³ as acylating agent for compound 1 under Ragnarsson's conditions gave rather discouraging results. The lower reactivity of the latter reagent, compared with the di-tert-butyl pyrocarbonate and its limited stability in the presence of 4-dimethylaminopyridine, could explain this observation already reported with dimethyl pyrocarbonate.⁹

Nevertheless the reaction of compound 1 with the mixed allyl *tert*-butyl pyrocarbonate 5, prepared by condensation of allyl chloroformate with sodium *tert*-butyl carbonate, allowed the synthesis of 1,3,4,6-tetra-O-acetyl-2-deoxy-2-diallyloxycarbonylamino- β -Dglucopyranose 6 in good yield (81 %). As expected, the attack on the other carbonyl function of compound 5 was disfavored for steric reasons; the compound 3 was nevertheless formed as a by-product (13 % yield).

The syntheses of other N,N-diacyl derivatives of D-glucosamine were also attempted in the aim of obtaining new glycosyl donors. The N-allyloxycarbonyl derivative 1 could not be benzoylated contrariwise to the analogous 2-acetamido compound 2 which could be transformed into the 2-acetamido-1,3,4,6-tetra-O-acetyl-N-benzoyl-2-deoxy- β -D-glucopyranose 7⁶ in 65 % yield by direct benzoylation with benzoyl chloride in pyridine.











13 R = COOAll**14** R = H**19** R = COPh

Nevertheless, the preparation of a N-acetyl-N-allyloxycarbonyl compound 8 was realized by N-acetylation of the compound 1, using an *in situ* formed mixed anhydride. The first attempt at acetylation of compound 1 using a mixed anhydride prepared from acetic acid, isobutyl chloroformate and tributylamine¹⁴ was negative and did not afford the expected compound 8. Another mode of preparation of mixed anhydrides was then considered. As already described by Pozdnev,¹⁵ dialkyl pyrocarbonates could be used for the

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TABLE 1. ¹H NMR chemical shifts of N-acylated and N-alkoxylated derivatives of D-glucosamine ^a

| | Solvent | T(°C) | Н-1 | Н-2 | Н-3 | H-4 | Н-5 | H6a | Нбb | NAC | % rotamer | |
|------------|----------|-------|------|--------------|------|------|--------------|------|------|------|-----------|--|
| 1 | CD3COCD3 | 25 | 5.86 | 3.85 | 5.33 | 5.04 | 3.94 | 4.27 | 4.08 | | | |
| • | CD3COCD3 | 25 | 6.36 | 4.37 | 5.77 | 5.07 | 3.99 | 4.29 | 4.17 | | | |
| 4b | CD3SOCD3 | 25 | 6.38 | 4.31 | 5.71 | 4.95 | 3.99 | 4.20 | 4.02 | 2.18 | 0.40 | |
| 4 | CD3SOCD3 | 60 | 6.30 | 4.83 4.62 | 5.69 | 4.95 | 3.95 | 4.20 | 4.08 | 2.30 | 0.00 | |
| 9 | CD3COCD3 | 25 | 6.37 | 4.45 | 5.78 | 5.09 | 3.99 | 4.30 | 4.08 | | | |
| ٢ | CD3SOCD3 | 6 | 6.40 | 4.34 | 5.78 | 4.94 | 4.02 | 4.20 | 4.06 | | | |
| 8 b | CD3SOCD3 | 25 | 6.38 | 4.30 | 5.71 | 4.97 | 4.02 | 4.21 | 4.01 | 2.25 | 0.45 | |
| | CD3SOCD3 | 60 | 6.29 | 4.60 | 5.68 | 4.97 | 4.02 3.99 | 4.2 | 4.01 | 2.30 | CC.N | |
| 6 | CD3COCD3 | 25 | 6.48 | 4.58 | 5.87 | 5.16 | 4.07 | 4.34 | 4.12 | | | |
| 10 | CD3COCD3 | 25 | 5.24 | 4.27 | 5.68 | 5.02 | 3.85 | 4.28 | 4.10 | | | |
| 11 | CD3COCD3 | 25 | 5.30 | 4.23 | 5.69 | 5.00 | 3.85 | 4.28 | 4.08 | | | |

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TABLE I (continued).

| 12 CD3COCD3 25 5.36 4.30 5.70 5.03 3.93.3.83 4.28 4.11 15 ^b CD3COCD3 25 5.36 4.30 5.01 4.87 3.93.3.83 4.28 4.11 15 ^b CD3SOCD3 25 5.29 4.09 5.61 4.87 3.86 4.19 4.07 2.28 0.43 15 ^b CD3SOCD3 25 5.29 4.09 5.61 4.87 3.86 4.19 4.07 2.28 0.43 16 ^b CD3SOCD3 25 5.29 4.90 5.61 4.87 3.86 4.19 4.07 2.36 0.57 16 ^b CD3SOCD3 25 5.28 4.86 3.85 4.21 4.07 2.35 0.53 16 ^b CD3SOCD3 90 5.21 4.46 5.58 4.86 3.85 4.19 4.07 2.35 0.53 17 CD3SOCD3 90 5.21 4.86 3.85 4.19 | | Solvent | T(°C) | I-H | Н-2 | Н-3 | H-4 | H-5 | H6a | Нбb | NAC | % rotamer |
|--|-----------------|-----------------------------------|-------|---------------------------|---------------------------|---------------------------|---------------------------|---------------------------|--------------------------------|---------------------------|------|-----------|
| 15 ^b CD3SOCD3 25 5.29 4.09 5.61 4.87 3.86 4.19 4.07 2.28 0.43 16 ^b CD3SOCD3 90 5.18 4.42 5.57 4.88 3.85 4.19 4.07 2.36 0.57 16 ^b CD3SOCD3 25 5.29 4.08 5.57 4.88 3.85 4.19 4.07 2.36 0.57 16 ^b CD3SOCD3 25 5.53 4.89 3.85 4.19 4.07 2.36 0.57 16 ^b CD3SOCD3 90 5.20 4.40 5.53 4.89 3.85 4.19 4.07 2.35 0.57 17 CD3SOCD3 90 5.21 4.59 5.58 4.89 3.85 4.19 4.07 2.35 0.53 17 CD3SOCD3 90 5.21 4.89 3.89 4.19 4.07 2.35 0.53 18 CD3SOCD3 90 5.10 4.15 3.80< | 12 | CD3COCD3 | 25 | 5.36 5.44 ^c | 4.30 4.32 ^c | 5.70 4.59c | 5.03 4.21c | 3.93-3.83 3.93-3.83 | 4.28 3.93-3.83 ^c | 4.11 3.66 ^c | | |
| CD3SOCD3 90 5.18 4.42 5.57 4.88 3.83 4.19 4.07 2.33 0.47 16 ^b CD3SOCD3 25 5.28 4.08 5.63 4.86 3.85 4.21 4.00 2.27 0.47 16 ^b CD3SOCD3 25 5.28 4.08 5.63 4.86 3.85 4.21 4.00 2.27 0.47 17 CD3SOCD3 90 5.21 4.40 5.58 4.86 3.85 4.19 4.00 2.35 0.53 17 CD3SOCD3 90 5.21 4.59 5.59 4.86 3.85 4.19 4.07 2.35 0.53 17 CD3SOCD3 90 5.21 4.56 4.86 3.86 3.80 4.19 4.07 2.35 0.53 17 CD3SOCD3 90 5.21 4.56 4.89 3.80 4.19 4.07 2.35 0.53 18 CD3SOCD3 25 6.18 4.15 3.80 4.19 4.07 2.35 0.53 2.35 0.53 | 15 b | CD3SOCD3 | 25 | 5.29 | 4.09 1.67 | 5.61 | 4.87 | 3.86 3.86 | 4.19 4.10 | 4.07 | 2.28 | 0.43 |
| 16 ^b CD ₃ SOCD ₃ 25 5.28 4.08 5.63 4.86 3.85 4.21 4.00 2.27 0.47 CD ₃ SOCD ₃ 90 5.13 4.59 5.52 4.89 3.85 4.19 4.00 2.27 0.47 17 CD ₃ SOCD ₃ 90 5.21 4.40 5.58 4.89 3.85 4.19 4.00 2.35 0.53 17 CD ₃ SOCD ₃ 90 5.21 4.52 5.59 4.89 3.80 4.19 4.07 2.35 0.53 18 CD ₃ SOCD ₃ 90 5.21 4.55 4.15 3.80 4.19 2.35 0.53 18 CD ₃ SOCD ₃ 25 6.18 4.15 3.80 3.85 3.52 3.55 3.55 20 CD ₃ SOCD ₃ 25 6.18 4.15 5.30 3.17 4.17 4.07 2.35 0.53 20 CD ₃ SOCD ₃ 25 6.18 4.15 3.65 3.55< | | CD3SOCD3 | 90 | 5.18 | 4.42 | 5.57 | 4.88 | 3.83 | 4.19 | 4.07 | 2.33 | 10.0 |
| CD3SOCD3 90 5.13 4.17 5.58 4.86 3.82 4.19 4.04 2.31 4.04 2.31 4.04 2.31 4.04 2.31 4.04 2.31 4.04 2.31 4.04 2.31 4.04 2.31 4.04 2.31 4.04 2.31 4.04 2.31 4.04 2.31 4.04 2.31 4.04 2.31 4.04 2.35 4.05 2.35 4.07 2.35 4.07 2.35 4.07 2.35 4.07 2.35 4.03 2.35 3.05 3.55 3.05 3.55 3.05 3.55 3.05 3.05 3.05 3.05 3.05 | 16 ^b | CD3SOCD3 | 25 | 5.28 | 4.08 | 5.63 | 4.86 | 3.85 | 4.21 | 4.00 | 2.27 | 0.47 |
| 17 CD ₃ SOCD ₃ 90 5.21 4.52 5.59 4.89 3.80 4.19 4.07 2.35 18 CD ₃ COCD ₃ 25 6.18 4.15 5.13 4.17 4.24 4.05 20 CD ₃ COCD ₃ 25 6.18 4.15 5.13 4.17 4.24 4.05 20 CD ₃ COCD ₃ 25 6.18 4.15 5.13 4.17 4.24 4.05 | | CD ₃ SOCD ₃ | 90 | 5.20 | 4.40 | 5.58 | 4.89 4.86 | 3.82 | 4.19 | 4.00 4.04 | 2.31 | cc.0 |
| 18 CD3COCD3 25 6.18 4.15 5.30 5.13 4.17 4.24 4.05 20 CD3COCD3 25 6.50 4.31 5.52 5.12 4.08 4.31 4.29 | 17 | CD3SOCD3 | 06 | 5.21 5.40 ^c | 4.52 4.29c | 5.59 4.55 ^c | 4.89 4.15 ^c | 3.80 3.80 ^c | 4.19 3.85 ^c | 4.07 3.52 ^c | 2.35 | |
| 20 CD ₃ COCD ₃ 25 6.50 4.31 5.52 5.12 4.08 4.31 4.29 | 18 | CD3COCD3 | 25 | 6.18 | 4.15 | 5.30 | 5.13 | 4.17 | 4.24 | 4.05 | | |
| | 20 | CD3COCD3 | 25 | 6.50 | 4.31 | 5.52 | 5.12 | 4.08 | 4.31 | 4.29 | | |

a. Chemical shifts in ppm from internal TMS
b. Two rotamers were observed
c. Galactopyranose moiety

| | Simidnon WIMM H- | | r-acylaicu ailu /v· | aikuxyiaicu ucii | Values of D-BI | ucosamme, in o | euteroacetone. | | | |
|------------|-----------------------------------|-------|---------------------|------------------|----------------|------------------|----------------|------------------|-------------------|--|
| Product | Solvent | T(°C) | J1,2 | J2,3 | J3,4 | J4,5 | J5,6a | J5a6a | J6a,6b | |
| 1 | CD3COCD3 | 25 | 8.8 | 10.5 | 9.4 | 10 | 4.8 | 2.4 | 12.5 | |
| e | CD3COCD3 | 25 | 8.6 | 10.6 | 9.2 | 10.2 | 4.6 | 2.1 | 12.4 | |
| 4b | CD ₃ SOCD ₃ | 25 | 7.9 | 9.7 | 8.6 | 10.2 | 4.5 | 2.1 | 12.6 | |
| |) | | 8.5 | 10.8 | 9.3 | 10.0 | 4.5 | 2.1 | 12.6 | |
| 4 | CD ₃ SOCD ₃ | 6 | 8.5 | 10.6 | 8.9 | 10.1 | 4.7 | 2.5 | 12.5 | |
| 9 | CD3COCD3 | 25 | 8.5 | 10.7 | 9.1 | 10.1 | 4.5 | 2.1 | 12.5 | |
| ٢ | CD ₃ SOCD ₃ | 90 | 8.5 | 10.5 | 9.0 | 10.1 | 5.0 | 2.5 | 12.5 | |
| 8 b | CD ₃ SOCD ₃ | 25 | 7.9 | 9.8 | 8.5 | 10.2 | 4.3 | 2.1 | 12.6 | |
| | I | | 8.5 | 10.7 | 9.4 | 10.0 | 4.3 | 2.1 | 12.6 | |
| 9 0 | CD3SOCD3 | 90 | 8.3 | 10.5 | 0.6 | 10.1 | 4.7 | 2.5 | 12.4 | |
| 6 | CDC13 | 25 | 8.7 | 10.6 | 9.2 | 9.7 | 4.4 | 2.1 | 12.3 | |
| 10 | CD3COCD3 | 25 | 8.1 | 10.8 | 9.1 | 10.1 | 4.6 | 2.4 | 12.2 | |
| 11 | CD3COCD3 | 25 | 8.1 | 10.8 | 9.0 | 10.2 | 5.0 | 2.4 | 12.2 | |
| 12 | CD3COCD3 | 25 | 8.1 | 10.7 | 9.0 | 10.1 | 4.0 | 2.2 | 12.2 | |
| | | | 4.9C | 2.2 ^c | 7.9c | 1.3 ^c | ſ | 8.5 ^c | 12.5 ^c | |

5 ոք որ . dated date. and M aller 1----LE M 3 1 TABLE II ¹H NMR complim

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TABLE II (continued).

| Product | Solvent | T(°C) | J1,2 | J2,3 | J3,4 | J4,5 | J5,6a | J 5a6a | J6a,6b |
|-----------------|-----------------------------------|-------|------------|------------------|------------------|------------------|------------------|--------|-------------------|
| 4 v | | ų | ¢ | ¢ - | 0 | 001 | C 7 | 6 | |
| el. | ന്നുന്നു | G | 8.U 7.9 | 0.11 | 8.9 8.9 | 10.01 | 4 4 7 2 | 2.0 | 12.4 |
| 15 | CD ₃ SOCD ₃ | 6 | 8.5 | 11.0 | 9.0 | 10.1 | 5.0 | 2.7 | 12.2 |
| 16 ^b | CD ₃ SOCD ₃ | 25 | 8.4 | 11.2 | 8.8 | 10.0 | • | , | • |
| | 1 | | 8.0 | 11.0 | 8.9 | 10.0 | , | • | |
| 16 | CD3SOCD3 | 90 | 8.0 | 1.11 | 9.0 | 10.0 | 5.1 | 2.5 | 12.2 |
| 17 | CD ₃ SOCD ₃ | 90 | 8.0 | 11.3 | 8.9 | 10.0 | 4.7 | 2.6 | 12.2 |
| | • | | 4.9C | 2.4 ^c | 7.9 ^c | 1.8 ^c | 4.4 ^C | 6.9c | 10.7 ^c |
| 18 | CD3COCD3 | 25 | 3.4 | 9.8 | 9.5 | 10 | 4.2 | 1.8 | 12.1 |
| 20 | cD3cocD3 | 25 | 3.6 | 10.9 | 9.5 | 10.1 | 4.1 | • | 12.3 |
| | | | | | | | | | |

Coupling constants in Herz. Two rotamers were observed. Galactopyranose moiety.

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TABLE III. 13C NMR Chemical shifts of N-acylated and N-alkoxylated derivatives of D-glucosamine, in deuteroacetone at room temperature

| | C-1 | C-2 | C-3 | C-4 | C-5 | C-6 | NCOOR | NCOR | NCOCH ₃ |
|----------|----------------|----------------|----------------|----------------|----------------|----------------|------------------|------------------|--------------------|
| 1 | 93.19 | 55.80 | 73.27 | 69.36 | 73.39 | 62.58 | 156.61 | - | |
| 3 | 91.83 | 60.39 | 70.98 | 69.72 | 73.26 | 62.39 | ind. | | |
| 4b | 91.54 92.13 | 61.10 56.11 | 70.77 71.67 | 69.55 69.55 | 73.12 73.34 | 62.39 62.39 | 154.14 152.37 | 174.18 173.40 | 27.25 26.40 |
| 9 | 91.72 | 60.78 | 70.88 | 69.52 | 73.22 | 62.34 | 154.27,154.27 | | |
| 7 | 91.72 | 61.84 | 70.76 | 69.85 | 73.26 | 62.39 | ind. | 174.96 | 27.07 |
| 4 | 91.37 92.08 | 61.06 56.40 | 70.78 71.43 | 69.18 69.18 | 73.16 73.16 | 62.06 62.36 | 155.02 153.78 | 174.02 173.21 | 27.22 26.41 |
| 6 | 91.78 | 59.62 | 70.71 | 69.76 | 73.33 | 62.36 | ind. | 172.79 | |
| 10 | 100.12 | 61.85 | 71.20 | 70.30 | 72.34 | 62.73 | 154.43, 153.72 | | |
| 11 | 99.10 | 62.03 | 71.19 | 70.36 | 72.28 | 62.85 | 154.45, 153.70 | | |

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TABLE III (continued).

| | C-1 | C-2 | C-3 | C 4 | C-5 | C-6 | NCOOR | NCOR | NCOCH ₃ |
|-----------------|--|--------------------------------------|-----------------------------|--------------------------------------|--------------------------------------|--------------------------------------|------------------|------------------|--------------------|
| 12 | 101.00 97.00c | 61.96 71.46 ^c | 71.27 71.14 ^c | 70.25 67.65 ⁰ | 72.34 71.63 ^c | 62.77 69.14 ^c | 154.30, 153.65 | | |
| 15 ^b | 99.79 100.52 | 62.28 57.66 | 71.13 71.83 | 70.40 70.40 | 72.42 72.42 | 62.85 62.85 | 155.37 154.17 | 174.15 173.19 | 27.27 26.46 |
| 16b | 98.70 99.50 | 62.40 57.80 | 71.10 71.79 | 70.40 70.40 | 72.25 72.25 | 62.89 62.89 | 155.41 154.21 | 174.18 173.11 | 27.24 26.46 |
| 17b | 100.86 102.06 97.34 ^c | 62.21 57.18 71.67 ^c | 71.19 71.87 71.19c | 70.30 70.30 67.87 ^c | 72.30 72.30 71.59 ^c | 62.80 62.80 69.87 ^c | 155.02 153.94 | 174.10 173.41 | 27.48 26.81 |
| 18 | 91.20 | 53.76 | 70.50 | 69.16 | 71.56 | 62.44 | 156.73 | | |
| 20 | 92.11 | 54.00 | 70.87 | 69.41 | 71.53 | 62.46 | 156.85 | | |
| | | | | | | | | | |

Chemical shifts in ppm from internal TMS. Two rotamers were observed. Galatopyranose moiety.

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activation of carboxylic acids in the presence of a base. In particular, di-*tert*-butyl pyrocarbonate reacted with carboxylic acids in the presence of pyridine to afford a mixed anhydride of carboxylic and *tert*-butylcarbonic acids. Thus, treatment of a solution of compound **1**, with acetic acid (2.5 equiv), di-*tert*-butyl pyrocarbonate (5 eq) and a catalytic amount of 4-dimethylaminopyridine in acetonitrile afforded 2-acetamido-1,3,4,6tetra-O-acetyl-N-allyloxycarbonyl-2-deoxy- β -D-glucopyranose **8** in high yield (90 %). The use of a twofold excess of di-*tert*-butyl pyrocarbonate *versus* acetic acid was necessary to avoid the formation of acetic anhydride by reaction of acetic acid with the mixed anhydride formed in the reaction mixture.¹⁵

The similar reaction of the mixed anhydride of benzoic and *tert*-butylcarbonic acids (formed *in situ* by reaction of di-*tert*-butyl pyrocarbonate and benzoic acid) with compound 1 did not show any regioselectivity. A mixture of the *N*-allyloxycarbonyl-*N*-*tert*-butoxy-carbonyl derivative 3 and the expected *N*-allyloxycarbonyl-*N*-benzoyl compound 9 was thus obtained in respective yields of 46% and 45% due to the reaction of the *N*-allyloxy-carbonylamino function on both carbonyl groups of the mixed anhydride.

The structures of compounds **3**, **4**, **6**-**9** were established on the basis of ¹H and ¹³C NMR data (Tables I, II and III). *N*-Acetylation or *N*-alkoxycarbonylation of **1** resulted in a substantial deshielding of H-1, H-2 and H-3 (0.4 - 0.7 ppm, Table I). ¹H NMR spectra of **4**, **7** and **8** revealed the presence of two conformers, the proportions of which were dependent on their structures (**4** : 40/60; **7** : 75/25; **8** : 45/55). This observation could be due to restricted rotations around the amide bond. On increasing the temperature, a coalescence of signals appeared at 60 °C for H-1 and H-3 and at 90 °C for H-2 (FIG. 1). ¹³C NMR spectra of compounds **4** and **8**, also confirmed the presence of two rotamers at room temperature (chemical shift differences in the range 0.7-0.9 ppm for C-1 and C-3 and 4.6-5.0 ppm for C-2 between both conformations). In the case of the acetamido-*N*-benzoyl compound **7**, the ¹³C NMR spectrum presented broadened singlets for C-1, C-2 and C-3. Surprisingly the ¹H and ¹³C NMR spectra of the *N*-benzoyl carbamate **9** displayed one form only at room temperature, which could either indicate free rotation around the amide bond or the predominance (>95:5) of one conformer.

Afterwards, compounds 6 and 8 were used as donors in 1,2-*trans* glycosylation reactions, under the conditions reported earlier with the donor $1.^{1-4}$, ¹² The reactions were conducted with a slight excess of alcohol (1.1 equiv.) and one equivalent of trimethylsilyl trifluoromethanesulfonate, with respect to the β -acetate 6 or 8, at -30 °C, in dichloromethane.

Glycosylation of **6** with ethanol, 2-propanol and 1,2:3,4-di-O-isopropylidene- α -D-galactopyranose afforded the expected β -glycosides 10(81 %), 11(59 %) and 12(75 %) together with the N-allyloxycarbonyl oxazolidinone by-product 13 (16 %, 36 % and 22 %





yields, respectively). As we reported earlier,¹² the amount of the cyclization compound **13** increased with the decreasing reactivity of the alcohol. When *tert*-butyl alcohol was used as the acceptor, the latter derivative was obtained as the main product.

In the absence of alcohol, the reaction of 6 with trimethylsilyl trifluoromethanesulfonate afforded the oxazolidinone 13 in 89 % yield. Compound 13 could also be prepared by direct allyloxycarbonylation of the oxazolidinone 14^{16} with allyl chloroformate and 1,8-diazabicyclo[5,4,0]undec-7-ene (DBU) in dichloromethane.

Under the same conditions, glycosylation of **8** with the same acceptors (ethanol, 2propanol and 1,2:3,4-di-*O*-isopropylidene- α -D-galactopyranose) afforded the expected β glycosides **15**(68 %), **16**(56 %) or **17**(60 %), together with 1,3,4,6-tetra-*O*-acetyl-2-allyloxycarbonylamino-2-deoxy- α -D-glucopyranose **18**¹ (respective yields: 27 %, 30 % and 28 %). The formation of the α -acetate **18** was probably due to an intramolecular rearrangement, with $N \rightarrow O$ acetyl migration as already observed by Inch *et al.* with the 2-acetamido-*N*-benzoyl-2-deoxy-D-glucopyranose.^{6,7}

In order to confirm the latter assessment, compound **8** was treated with trimethylsilyl trifluoromethanesulfonate in the absence of alcohol, under the same conditions. A mixture of two compounds was thus recovered, the ¹H NMR spectrum of which exhibited the presence of an anomeric α -acetate for the first one (δ H-1 6.18 ppm, J_{1,2} = 3.4 Hz) and of a *N*-acetylallyloxycarbonylamino function for the second one (δ CH₃ = 2.40 ppm) in the ratio 3:2. Attempts to separate these two compounds were unsuccessful since the slower moving product rearranged to the faster one during column chromatography. We suggest that the slower moving product (tentatively assigned as 2-acetamido-3,4,6-tri-*O*acetyl-*N*-allyloxycarbonyl-2-deoxy-D-glucopyranose) could have been formed with the allyloxycarbonylamino function acting as the participating group. The faster moving derivative (identical with compound 18) could have been formed, contrariwise, *via* a *N*->O acetyl migration, with the acetamido function acting as the participating group.

The structure of the glycosides 10 - 12 and 15 - 17 were ascertained by ¹H and ¹³C NMR. The β -configurations were characterized by the values of $J_{1,2}$ and δ C-1 in ¹H and ¹³C NMR spectra respectively (Tables II and III). As reported for compounds 4, 7 and 8, the NMR spectra of compounds 15-17 also showed mixtures of conformers, at room temperature, in ratios depending on the solvent. When the temperature was raised to 90 °C, rotation around the amide bond became too fast for a spectroscopic differentiation of the rotamers (Tables I, II and III).

The β -acetate 9 was also tested as glycosyl donor under the same conditions. However, glycosylation with 2-propanol gave a complex mixture of compounds which was not analyzed. Furthermore, when the reaction was realized in the absence of alcohol, two major products were obtained in the mixture. Attempts to separate them were unsuccessful, but ¹H and ¹³C NMR spectra allowed assignment of the structure **19** to the minor one (17 % yield) and **20** to the major one (68 % yield). The formation of the latter could have resulted from a benzoyl migration from the nitrogen atom to the anomeric oxygen, as already observed starting from **8**.

The structures of compounds 19 and 20 were confirmed by chemical proofs. Thus, the benzoylation of the oxazolidinone 14 in pyridine afforded the *N*-benzoyl oxazolidinone 19 in 92 % yield, and the cleavage of the anomeric acetate of 1 with hydrazine acetate in dimethylformamide,¹⁷ followed by benzoylation of the free anomeric hydroxyl group gave a $6/1 \alpha/\beta$ anomeric mixture of 20 (yield 77 %) from which the α -anomer could be isolated in a pure form.

In conclusion, use of the mixed anhydrides or mixed pyrocarbonates allowed us to synthesize N-acyl-N-alkoxycarbonyl and N,N-dialkoxycarbonyl derivatives of D-glucosamine in high yields. These products could be used as donors in 1,2-*trans*-glycosylations, but cyclizations and rearrangements in the glycosylation conditions, gave by-products, limiting their interest as donors.

EXPERIMENTAL

General Procedures. Acetonitrile and dichloromethane were dried by refluxing respectively with calcium hydride and phosphorus pentoxide for six hours, then distilled and stored over 4 Å molecular sieves. Tetrahydrofuran was refluxed with sodiumbenzophenone mixture under a nitrogen atmosphere until a blue color was obtained, then distilled and stirred over 4 Å molecular sieves. Melting points were determined on an Electrothermal 9100 apparatus and were uncorrected. TLC analyses were performed on aluminium sheets coated with silica gel 60 F 254 Merck. Compounds were visualized by UV light or by spraying the TLC plates with dilute 15 % aqueous sulfuric acid, followed by charring at 150 °C for a few minutes. Column chromatography was performed on Merck-Kieselgel (230-400 mesh ASTM). Optical rotations were measured on a Perkin Elmer 241 polarimeter in a 1 dm cell. ¹H and ¹³C NMR spectra were recorded with a Brucker AM-300 spectrometer at 300 or 75.5 MHz respectively with tetramethylsilane as internal standard. Elemental analyses were realized by the "Laboratoire Central d'Analyses du CNRS" (Solaize, France).

Compounds 1 and 2 were prepared by published methods.^{1,18}

1,3,4,6-Tetra-O-acetyl-2-allyloxycarbonylamino-N-tert-butoxycarbonyl-2-deoxy- β -D-glucopyranose (3). A solution of 1,3,4,6-tetra-O-acetyl-2-allyloxycarbonylamino-2-deoxy- β -D-glucopyranose (1) (0.432 g, 1 mmol) in dry acetonitrile (5 mL) was treated with 4-dimethylaminopyridine (0.012 g, 0.1 mmol) and di-*tert*-butyl pyrocarbonate (0.262 g, 1.2 mmol) at room temperature overnight. The solution was then concentrated *in vacuo* and the residue was applied to the top of a silica gel column and then eluted with ethyl acetate/hexane (1:1, v/v) to give the pure compound 3 (0.463 g, 87 %) as a colorless oil: R_f 0.63 (ethyl acetate/hexane 1:1, v/v); $[\alpha]_D$ +24.6 (c 1.0, chloroform); ¹H and ¹³ C NMR, Tables I, II and III.

Anal. Calcd for C₂₃H₃₃NO₁₃ (531.50): C, 51.97; H, 6.26; N, 2.64. Found: C, 51.89; H, 6.27; N, 2.68.

2-Acetamido-1,3,4,6-tetra-O-acetyl-N-tert-butoxycarbonyl-2-deoxy- β -D-glucopyranose (4). Compound 4 was obtained from 2-acetamido-1,3,4,6-tetra-O-acetyl-2-deoxy- β -D-glucopyranose (2) as described for the preparation of 3 from 1. The crude product was purified by column chromatography (ethyl acetate/hexane 2:1, v/v) to give 4 as a colorless oil (0.420 g, 85 %): R_f 0.68 (ethyl acetate/hexane 1:1, v/v); [α]_D +14.8 (c 1.0, chloroform); ¹H and ¹³C NMR, Tables I, II and III.

Anal. Calcd for C₂₁H₃₁NO₁₂ (489.46): C, 51.53; H, 6.38; N, 2.86. Found: C, 51.39; H, 6.41; N, 2.86.

Allyl tert-Butyl Pyrocarbonate (5). To a 100 mL, three-necked flask, fitted with a mechanical stirrer, a 50 mL pressure equalizing dropping funnel and a gas-inlet tube (6 mm internal diameter) extending nearly to the bottom of the flask, was poured a solution of sodium tert-butoxide (3.8 g, 39.6 mmol) in dry tetrahydrofuran (40 mL), under nitrogen. The reaction flask was immersed in an ice-salt bath maintained at -15 °C, and a stream of anhydrous carbon dioxide was passed through the cold solution for 0.5 h, with stirring. The formation of a thick slurry in the reaction flask was thus observed. Allyl chloroformate (4.6 mL, 43.3 mmol) was then added dropwise followed by tetrahydrofuran (15 mL). When the addition was complete, the solution was allowed to reach room temperature and stirring was maintained for 3 h. The solution was concentrated in vacuo, the residue was dissolved in dichloromethane (100 mL) and the organic phase washed twice with water, dried and concentrated to dryness. Finally, the obtained residue was centrifugated to afford a crude colorless syrup 5 which was directly used for the further step (4.8 g, 60 % yield). ¹H NMR (CDCl₃) δ 5.95 (m, 1H, CH=), 5.47-5.30 (m, 2H, $CH_{2}=$), 4.78-4.70 (m, 2H, allyl - CH_{2}), 1.57 (s, 9H, 3 CH_{3}); ¹³C NMR (CDCl₃) δ 148.92, 148.29 (2 C=O), 130.43 (=CH-), 120.00 (CH₂=), 85.94 (C(CH₃)₃), 69.73 (allyl CH2-), 27.39 (C(CH3)3).

1,3,4,6-Tetra-O-acetyl-2-deoxy-2-diallyloxycarbonylamino- β -D-glucopyranose (6). Compound 6 was obtained from 1 (1.078 g, 2.5 mmol) and 5 (0.758 g, 3.75 mmol) as already described for the preparation of 3 from 1. The crude product was purified by column chromatography (ethyl acetate/hexane 1:1 v/v) to afford successively the by-product **3** (0.173 g, 13 % yield) and the expected product **6** as a colorless oil (1.044 g, 81 % yield): Rf 0.58 (ethyl acetate/hexane 1:1 v/v); $[\alpha]_D$ +16.2 (c 1.8, chloroform); ¹H and ¹³C NMR, Tables I, II and III.

Anal. Calcd for C₂₂H₂₉NO₁₃ (515.46): C, 51.26; H, 5.67; N, 2.72. Found: C, 51.04; H, 5.83; N, 2.74.

2-Acetamido-1,3,4,6-tetra-O-acetyl-N-benzoyl-2-deoxy- β -D-glucopyranose (7). A solution of 2 (1g, 2.57 mmol) and benzoyl chloride (0.3 mL, 0.363 g, 2.56 mmol) in pyridine (10 mL) was stirred overnight at room temperature. After concentration *in vacuo*, the crude residue was directly applied to a silica gel column (ethyl acetate/hexane 3: 2, v/v) to afford the chromatographically pure product 7 as an amorphous solid (0.799 g, 63 %): R_f 0.40 (ethyl acetate/hexane 1:1, v/v); [α]_D -22.0 (c 1.0, chloroform) (lit.⁶ [α]_D - 22; c 0.85, chloroform); ¹H and ¹³C NMR Tables I, II and III.

2-Acetamido-1,3,4,6-tetra-O-acetyl-N-allyloxycarbonyl-2-deoxy- β -Dglucopyranose (8). Acetic acid (0.297 mL, 5.2 mmol) was added dropwise to a cold (-10 °C) solution of di-*tert*-butyl pyrocarbonate (2.27 g, 10.4 mmol) in dry acetonitrile (10 mL), followed by a catalytic amount of 4-dimethylaminopyridine (0.020 g, 0.16 mmol). The mixture was stirred for 0.5 h at -10 °C and a solution of 1 (0.897 g, 2.08 mmol) in acetonitrile (5 mL) was added in once. The reaction mixture was allowed to reach room temperature and stirring was continued overnight. After concentration *in vacuo*, the brown residue was directly applied to a column of silica gel (ethyl acetate/hexane 1 :1 v/v) to give the pure compound 8 (0.886 g, 90 % yield) as a colorless oil which crystallized from ethanol and hexane: mp 99 °C; R_f 0.55 (ethyl acetate/hexane 1:1 v/v); $[\alpha]_D$ +9.4 (*c* 1.0, chloroform); ¹H and ¹³C NMR, Tables I, II and III.

Anal. Calcd for C₂₀H₂₇NO₁₂ (473.42): C, 50.74; H, 5.75; N, 2.96. Found: C, 50.53; H, 5.66; N, 3.08.

1,3,4,6-Tetra-O-acetyl-2-allyloxycarbonylamino-N-benzoyl-2-deoxy- β -D-glucopyranose (9). Benzoic acid (0.767 g, 6.28 mmol) was added to a cold (-10 °C) solution of di-*tert*-butyl pyrocarbonate (1.83 g, 8.38 mmol) in dry acetonitrile (10 mL) followed by a catalytic amount of 4-dimethylaminopyridine. The mixture was stirred for 0.5 h at -10 °C and a solution of 1 (1.083 g, 2.51 mmol) in acetonitrile was added in once. After the same work-up procedure as precedingly, the crude product was purified by column chromatography (ethyl acetate/hexane 2:3 v/v) to afford the pure compounds 3 (0.613 g, 46 %) and 9 (0.605 g, 45 %). Compound 9 was recrystallized from ethanol: mp 130 °C; R_f 0.65 (ethyl acetate/hexane 1:2, v/v); $[\alpha]_D$ +22.3 (c 1.0, chloroform); ¹H and ¹³C NMR, Tables I, II and III.

Anal. Calcd for C₂₅H₂₉NO₁₂ (535.49): C, 56.07; H, 5.46; N, 2.62. Found: C, 56.22; H, 5.36; N, 2.68.

General procedure for glycosylation reactions. The β -acetate 6 or 8 (0.5 mmol) and the acceptor alcohol (0.55 mmol) were added to dry alcohol-free dichloromethane (25 mL). The reaction mixture was flushed with nitrogen while cooling to -30 °C. Trimethylsilyl trifluoromethanesulfonate (FLUKA) (95.2 μ L, 0.525 mmol) was then introduced through a syringe and the nitrogen flush maintained for 0.5 h. The mixture was stirred for 16 h at - 30 °C and the reaction was quenched by addition of triethylamine (0.148 mL, 1.05 mmol). The mixture was then poured into a saturated aqueous solution of sodium hydrogencarbonate and the products were extracted with dichloromethane. After drying and concentration *in vacuo*, the organic phase led to a mixture which was purified by column chromatography.

Ethyl 3,4,6-Tri-O-acetyl-2-deoxy-2-diallyloxycarbonylamino- β -Dglucopyranoside (10). Prepared following the general glycosylation procedure (6 as donor, ethanol as acceptor alcohol). Purification of the residue on a silica gel column (ethyl acetate/hexane 1:1, v/v) afforded compound 10 (0.203 g, 81 %) as a colorless oil: R_f 0.68 (ethyl acetate/hexane 1:1, v/v); $[\alpha]_D$ -9.5 (c 1.0, chloroform); ¹H and ¹³C NMR, Tables I, II and III.

Anal. Calcd for C₂₂H₃₁NO₁₂ (501.47): C, 52.69; H, 6.23; N, 2.79. Found: C, 52.74; H, 6.25; N, 2.87.

Further elution afforded compound 13 (0.037 g, 16 %), Rf 0.38.

Isopropyl 3,4,6-Tri-O-acetyl-2-deoxy-2-diallyloxycarbonylamino- β -D-glucopyranoside (11). Prepared following the general glycosylation procedure (6 as donor, 2-propanol as acceptor alcohol). Purification of the residue on a silica gel column afforded compound 11 (0.152 g, 59 %) as a colorless oil: R_f 0.63 (ethyl acetate/hexane 1:1, v/v); [α]_D-18.8 (c 1.0, chloroform); ¹H and ¹³C NMR, Tables I, II and III.

Anal. Calcd for C₂₃H₃₃NO₁₂ (515.50): C, 53.58; H, 6.45; N, 2.72. Found: C, 53.38; H, 6.50; N, 2.68.

Further elution afforded compound 13 (0.075 g; 36 %).

6-O-(3,4,6-Tri-O-acetyl-2-deoxy-2-diallyloxycarbonylamino-β-D-glucopyranosyl)-1,2:3,4-di-O-isopropylidene-α-D-galactopyranose (12). Prepared following the general glycosylation procedure (6 as donor, 1,2:3,4-di-O-isopropylidene-α-D-galactopyranose as acceptor alcohol). Purification on a column of silica gel afforded compound 12 (0.270 g, 75 %) as a colorless oil: R_f 0.59 (ethyl acetate/hexane 1:1, v/v); [α]_D-44.5 (c 1.0, chloroform); ¹H and ¹³C NMR, Tables I, II and III.

Anal. Calcd for C₃₂H₄₅NO₁₇ (715.69): C, 53.70; H, 6.34; N, 1.96. Found: C, 53.98; H, 6.39; N, 2.02.

Further elution afforded a mixture of compound 13 (0.031 g; 15 %) and unreacted acceptor alcohol (0.033 g, 22 %).

3-Allyloxycarbonyl-[3,4,6-tri-O-acetyl-1,2-dideoxy-α-D-glucopyra-

no]-[2,1-d]-2-oxazolidinone (13). Method A : Compound 13 could be prepared from 6 as previously described for glycosylation reactions but in absence of alcohol, with a 89 % yield after column chromatography (ethyl acetate/hexane 1:1 v/v). The product 13 crystallized from dichloromethane-hexane and was recrystallized from ethanol: mp 106 °C (ethanol); R_f 0.38 (ethyl acetate/hexane 1:1, v/v); $[\alpha]_D$ -25.7 (*c* 1.0, chloroform); ¹H NMR (CD₃COCD₃) δ 6.13 (d, 1H, J_{1,2} = 7.1 Hz, H-1), 5.99 (m, 1H, -CH=), 5.55 (dd, 1H, J_{2,3} = 4.0 Hz, J_{3,4} = 4.2 Hz, H-3), 5.44 and 5.25 (m, 2H, CH₂=), 5.02 (ddd, 1H, J_{2,4} = 0.6Hz, J_{4,5} = 7.4 Hz, H-4), 4.79 - 4.68 (m, 2H, allyl -CH₂-), 4.67 (ddd, 1H, H-2), 4.27 (dd 1H, J_{5,6a} = 5.5 Hz, J_{6a,6b} = 12.2 Hz, H-6a), 4.22 (dd, 1H, J_{5,6b} = 3.6 Hz, H-6b), 4.08 (ddd, 1H, H-5), 2.06, 2.01, 1.99 (3s, 9H, 3CH₃CO). ¹³C NMR (CD₃COCD₃) δ 170.64, 169.74, 169.63 (3C, C=O acetates), 150.97, 150.18 (2C, N-C=O), 132.47 (CH=), 118.82 (CH₂=), 94.85 (C-1), 70.14, 68.50, 67.15 (C-3,4,5), 68.07 (CH₂-allyl), 63.79 (C-6), 55.14 (C-2), 20.76, 20.60, 20.58 (3C, CH₃ acetates).

Anal. Calcd for C₁₇H₂₁NO₁₁ (415.345): C, 49.16; H, 5.10; N, 3.37. Found: C, 48.98; H, 5.10; N, 3.50.

- Method B : A solution of the oxazolidinone 14^{12} (0.780 g, 2.3 mmol) in dry dichloromethane was stirred for 2 h with 1,8-diazabicyclo[5,4,0]undec-7-ene (DBU) (1.7 mL, 11.5 mmol, 5 eq) and allyl chloroformate (0.6 mL, 5.6 mmol). After concentration *in vacuo*, the crude residue was purified by column chromatography to give the pure product 13 (0.760 g) in 85 % yield.

Ethyl 2-Acetamido-3,4,6-tri-O-acetyl-N-allyloxycarbonyl-2-deoxy- β -D-glucopyranoside (15). Prepared following the general glycosylation procedure (8 as donor, ethanol as acceptor alcohol). Purification of the residue on a column of silica gel (ethyl acetate/hexane 1:1, v/v) afforded compound 15 as a colorless oil (0.156 g, 68 %): R_f 0.58 (ethyl acetate/hexane 1:1, v/v); [α]_D -17.0 (*c* 1.0, chloroform); ¹H and ¹³C NMR, Tables I, II and III.

Anal. Calcd for C₂₀H₂₉NO₁₁ (459.44): C, 52.28; H, 6.36; N, 3.05. Found: C, 52.31; H, 6.43; N, 3.10.

Further elution afforded compound 18 (0.058 g, 27 %).

Isopropyl 2-Acetamido-3,4,6-tri-*O***-acetyl-***N***-allyloxycarbonyl-2-deoxy-** β **-D-glucopyranoside (16)**. Obtained, following the general glycosylation procedure (8 as donor, 2-propanol as acceptor alcohol), in 56 % yield after chromatographic purification; 16 was a colorless oil: $R_f 0.73$ (ethyl acetate/hexane 1:1, v/v); $[\alpha]_D$ -23.8 (*c* 1.0, chloroform); ¹H and ¹³C NMR Tables I, II and III.

Anal. Calcd for C₂₁H₃₁NO₁₁ (473.46): C, 53.27; H, 6.60; N, 2.96. Found: C, 53.13; H, 6.55; N, 3.20.

Further elution afforded the compound 18 (0.065g, 30 %).

6-O-(2-Acetamido-3,4,6-tri-O-acetyl-N-allyloxycarbonyl-2-deoxy-β-D-glucopyranosyl)-1,2:3,4-di-O-isopropylidene-α-D-galactopyranose (17). Prepared as described above from 8 (0.50 mmol) and 1,2:3,4-di-O-isopropylidene-α-D-galactopyranose (0.55 mmol). Purification of the residue on a column of silica gel (ethyl acetate/hexane 1:2, v/v) afforded successively the disaccharide 17 (0.202 g, 60 %) and the α-acetate 18 (0.058 g, 27 %) followed by the unreacted alcohol (0.041 g, 28 %). Compound 17 was a colorless oil: R_f 0.58 (ethyl acetate/hexane 1:1, v/v); [α]D -43.7 (c 1.0, chloroform); ¹H and ¹³C NMR, Tables I, II and III.

Anal. Calcd for C₃₀H₄₃NO₁₄ (672.65): C, 53.48; H, 6.43; N, 2.08. Found: C, 53.22; H, 6.45; N, 2.05.

1,3,4,6-Tetra-O-acetyl-2-allyloxycarbonylamino-2-deoxy- α -D-glucopyranose (18). Compound 8 (0.50 mmol) was treated overnight with trimethylsilyl trifluoromethanesulfonate (0.50 mmol) at -30 °C in dichloromethane (25 mL). After usual work-up, TLC of the crude product showed the presence of two compounds (R_f 0.70 and R_f 0.52 in dichloromethane/acetone 6:1, v/v) and ¹H NMR exhibited an anomeric acetate (δ H-1 6.18 ppm) and a *N*-acetyl group (δ CH₃ ~ 2.40 ppm) in the ratio 3:2. Attempts to separate the two compounds on a column of silica gel with the same eluent afforded only the first product (R_f = 0.70) in 82 % yield, which crystallized from ether. Compound 18 had mp 120 °C (lit.¹ mp 119-121 °C); R_f 0.51 (ethyl acetate/hexane 1:1, v/v); [α]_D+91.7 (*c* 1.0, chloroform); ¹H and ¹³C NMR, Tables I, II and III.

3-Benzoyl-[3,4,6-tri-*O*-acetyl-1,2-dideoxy-α-D-glucopyrano]-[2,1-d]-**2-oxazolidinone (19).** To a cold (0 °C) solution of oxazolidinone 14¹² (0.200 g, 0.60 mmol) in dry pyridine was added benzoyl chloride (0.14 mL, 2 eq) and 4-dimethylaminopyridine (0.005 g). The solution was allowed to reach room temperature and stirring was maintained overnight. After concentration of the solution, the residue was applied to the top of a silica gel column (ethyl acetate/hexane 1:1, v/v) to give product 20 which crystallized from ethanol (0.240 g, 92 % yield): mp 162 °C; R_f 0.63 (ethyl acetate/hexane 1:1, v/v); $[\alpha]_D$ -69.3 (*c* 1.0, chloroform); ¹H NMR (CD₃COCD₃) δ 7.77-7.44 (m, 5H, H aromat.), 6.21 (d, 1H, J_{1,2} = 7.1 Hz, H-1), 5.55 (dd, 1H, J_{3,2} = 3.5 Hz, J_{3,4} = 2.6 Hz, H-3), 5.11 (m, 1H, J_{4,5} = 6.1 Hz, H-4), 4.98 (ddd, 1H,J_{2,4} = 1.2 Hz, H-2), 4.35-4.25 (m, 3H, H-5,6a,6b), 2.10, 2.04, 1.87 (3S,9 H, 3CH₃CO). ¹³C NMR (CD₃COCD₃) δ 170.69, 169.70, 169.68, 169.50 (4 *C*=O), 152.66 (N-*C*=O), 134.27 - 128.60 (C aromat.), 94.87 (C-1), 69.73, 67.54, 67.27 (3C, C-3,4,5), 64.18 (C-6), 53.85 (C-2), 20.73, 20.60, 20.60 (3C, CH₃ acetates).

Anal. Calcd for C₂₀H₂₁NO₁₀ (435.37): C, 55.17; H, 4.86; N, 3.22. Found: C, 55.04; H, 4.98; N, 3.14.

3,4,6-Tri-O-acetyl-2-allyloxycarbonylamino-1-O-benzoyl-2-deoxy- α -D-glucopyranose (20). The β -acetate 1 (1g, 2.32 mmmol) was treated for 5 min at 50 °C with hydrazine acetate (0.195 g, 2.78 mmol) in dry dimethylformamide. After addition of ethyl acetate, the mixture was washed with brine (2 x 5 mL), and the organic phase dried and concentrated *in vacuo*. The residue was quickly purified by column chromatography to afford the α , β -hemiacetal (0.768 g, 85 %) which was dissolved in pyridine (5 mL) and treated overnight with benzoyl chloride (0.46 mL, 2 eq). After concentration, the crude residue was purified by column chromatography (ethyl acetate/hexane 1:1, v/v) to afford successively the pure α -anomer 20 (0.390 g, 40 %) and an α , β -mixture (0.360 g, 37 %, α : β =7:3). Compound 20 α was an amorphous solid, R_f 0.60 (ethyl acetate/hexane 1:1, v/v); $[\alpha]_D$ +115.7 (c 1.0, chloroform); ¹H NMR and ¹³C NMR, Tables I, II and III.

Anal. Calcd for ₂₃H₂₇NO₁₁ (493.45): C, 55.98; H, 5.52; N, 2.84. Found: C, 55.63; H, 5.66; N, 2.77.

Compound 20 β had R_f 0.58 (ethyl acetate/hexane 1:1, v/v); ¹H NMR (CDCl₃) δ 6.00 (1H, H-1, J_{1,2} = 6.8 Hz).

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